

Bioinspired liquid crystal patterns to command living matter

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SC0019105

APS March Meeting 2020

**Session F18: Non-equilibrium bioinspired modes of assembled materials,
March 3, 2020**

Contributing graduate students, Chemical Physics Interdisciplinary Program



Shuang Zhou, PhD 2016;
now Assistant professor at
U Mass, Amherst



Chenhui Peng, PhD 2016;
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Greta Babakhanova,
PhD 2019, now a
researcher at NIST



Bingxiang Li, PhD 2019,
now a PDF at AMLCI



Runa Koizumi,
current PhD student



Taras Turiv,
current PhD student

Collaborators

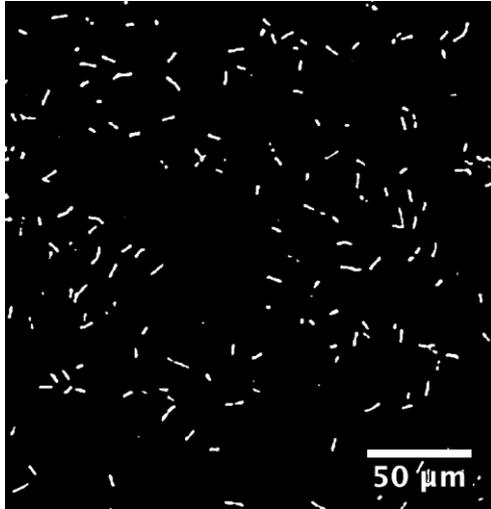
Kent State University:

- Dr. Sergij Shiyankovskii, AMLCI
- Dr. Hao Yu and Prof. Qi-Huo Wei, AMLCI, Physics and CPIP
- Miss Jess Krieger and Prof. Min-Ho Kim, Biological Sciences
- Mr. Robert Lastowski, REU student

Our colleagues:

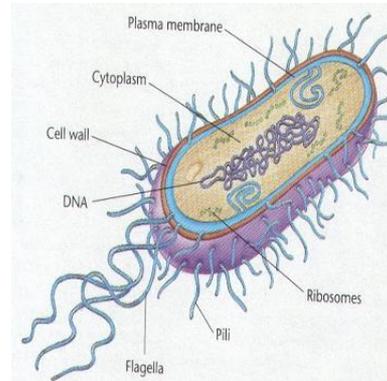
- Prof. Igor Aranson, Department of Biomedical Engineering, Pennsylvania State University
- Kristian Thijssen and Julia M. Yeomans, The Rudolf Peierls Centre for Theoretical Physics, Clarendon Laboratory, University of Oxford, UK
- Amin Doostmohammadi, University of Oxford, now at The Niels Bohr Institute, University of Copenhagen, Denmark
- Mikhail M. Genkin, Cold Spring Harbor Laboratory

Motivation No.1: Swimming bacteria: Can we extract a useful work from them?



Swimming Bacillus Subtilis

Effective swimmers at microscale, bacteria are “ready-to-use” for microtransport, delivery, mixing

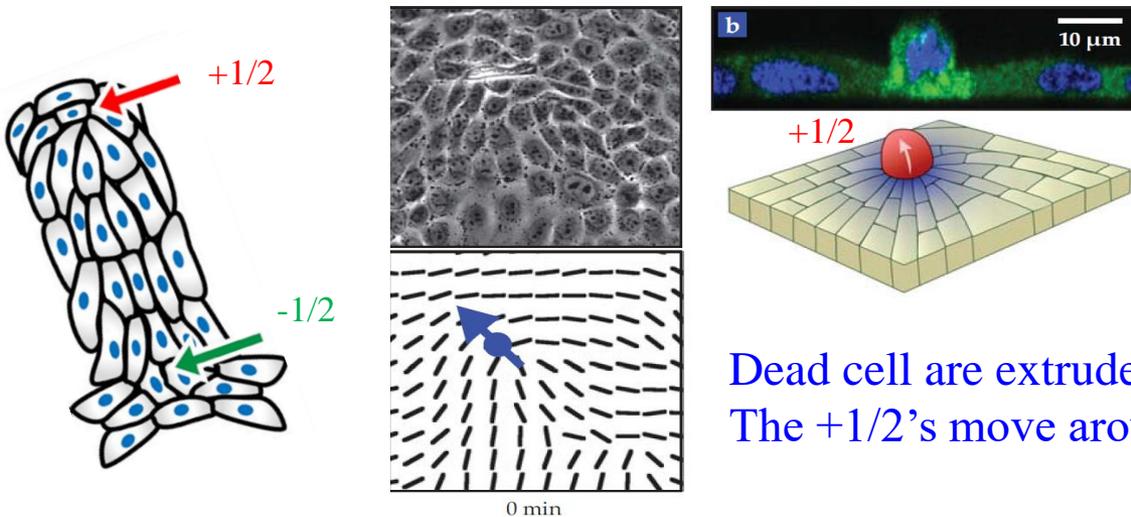


Can we force the swimming bacteria to deliver cargos, drugs, to run micromachines, to transform them into horses of microcosm?

Problem: Visual, audio and tactile communication channels are not available

Motivation No.2: Epithelium: Can we design it?

In epithelia, cells form an orientationally ordered nematic with topological defects, T.B. Saw et al, *Topological defects in epithelia govern cell death and extrusion*, Nature **544**, 212 (2017):



Dead cells are extruded at the $+1/2$ cores;
The $+1/2$'s move around in the plane of film

Topological defects are intrinsic to many other active matter systems

Sanchez et al, Nature **491**, 431 (2012), Zhou et al, PNAS 111, 1265 (2014)

We do not know *where* the defects would emerge nor *how to control* them
Can we develop a template to produce and pin the defects at predesigned locations?

Liquid crystals as guiding and templating medium

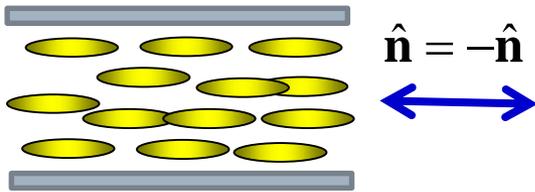
This work:

Replace an isotropic aqueous environment natural to many biological microsystems by an orientationally ordered liquid crystal;

Use patterned liquid crystals as a “communication channel” to command swimming bacteria and epithelium patterns

Why liquid crystals are different from water

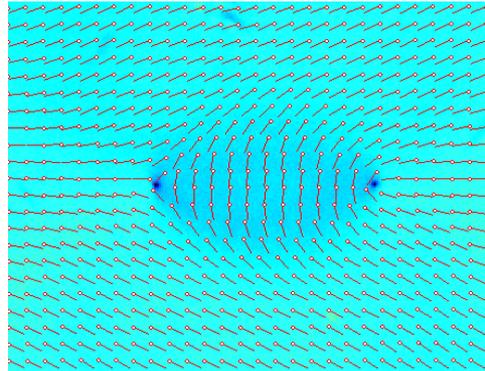
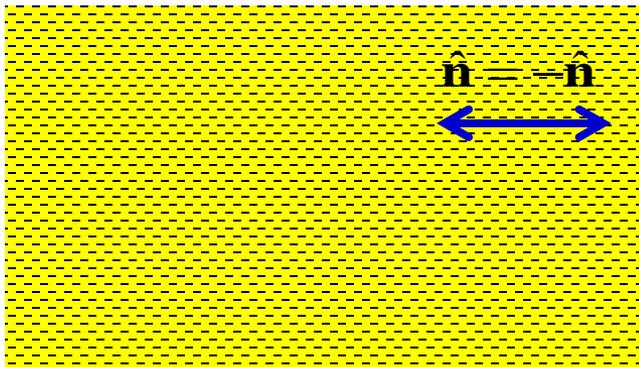
□ Anisotropy:



anisotropy axis of dielectric permittivity,
viscous drag, birefringence, etc.

□ Patterning capabilities

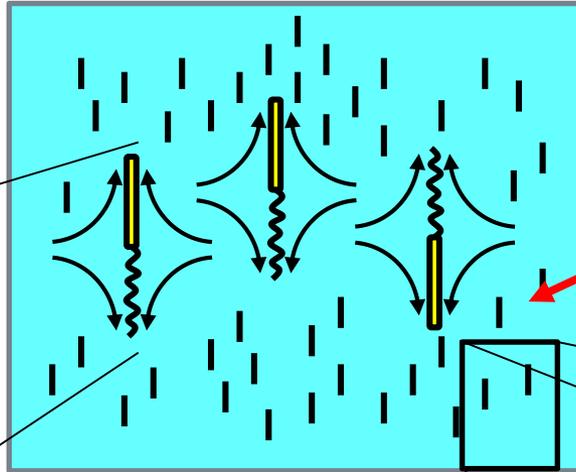
Can be prepared as uniformly aligned or as a pattern, using a photoalignment technique



Liquid Crystals to Command Swimming Bacteria

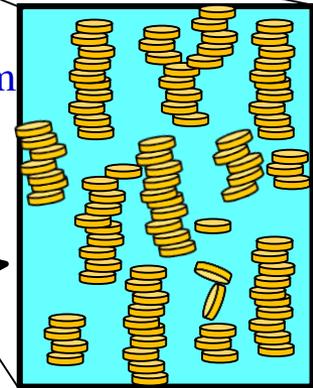
B. Subtilis, swims by rotating helical flagella; activity is controlled by oxygen/nitrogen supply

Body:
2-5 μm



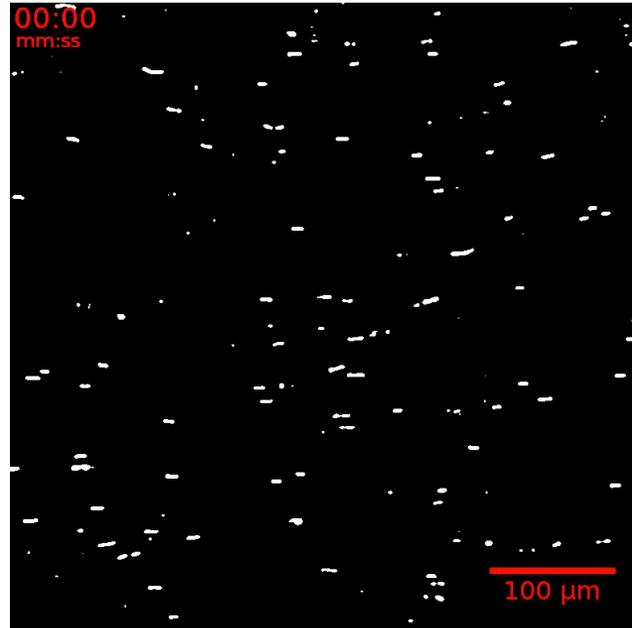
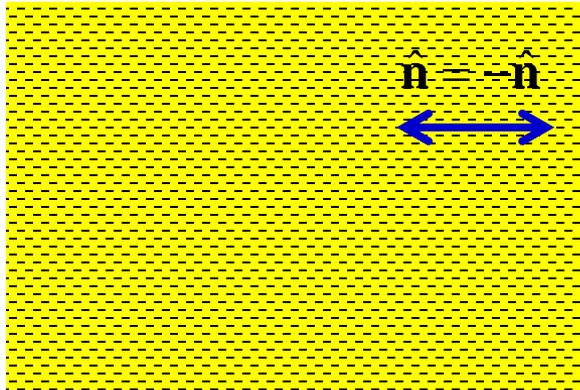
Chromonic LC

10-100 nm



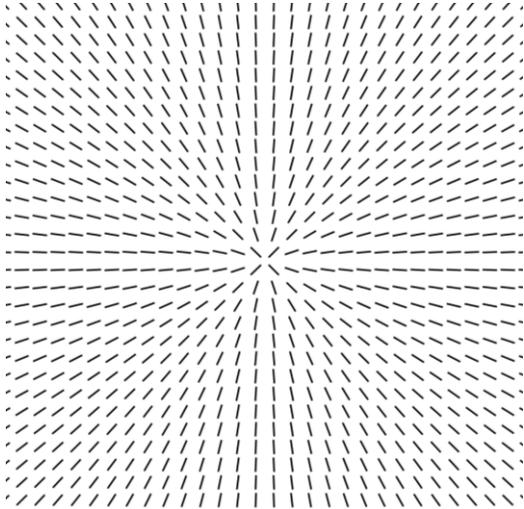
Main feature: an independent control of orientational order and activity

Uniform alignment: bacteria follow the director

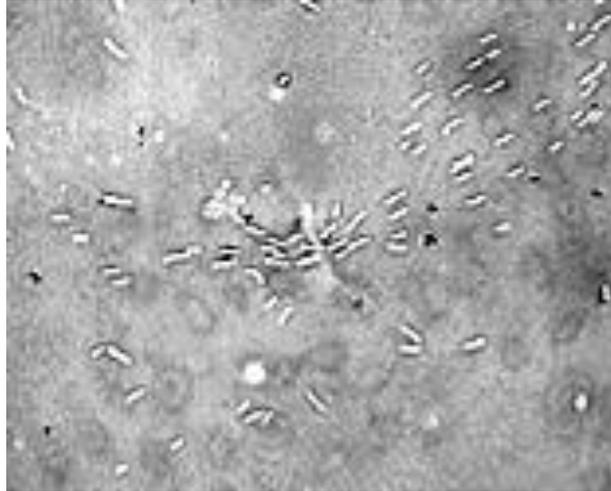


Swimming parallel to the director;
no distinction between right and left,
thus no useful work...

Pure splay: Bacteria swim || director; bipolar motion



$$\text{splay} = \hat{\mathbf{n}} \nabla \cdot \hat{\mathbf{n}} \neq 0$$

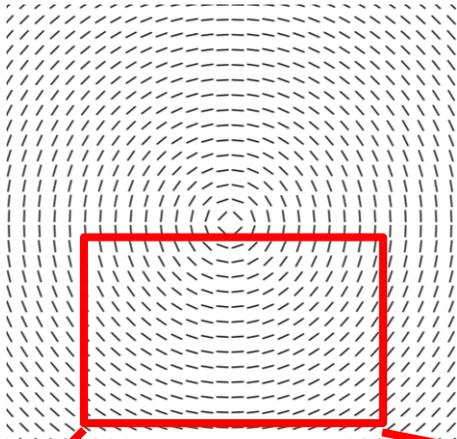


$$c_B = 10^{14} / \text{m}^3$$

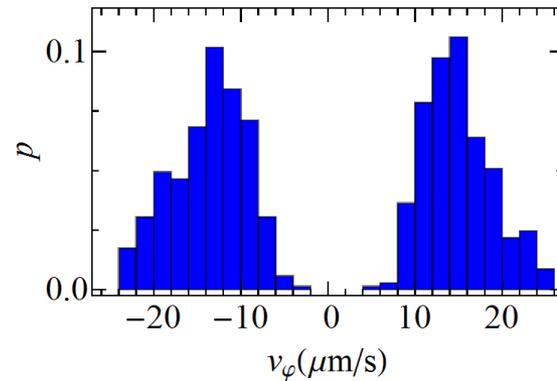
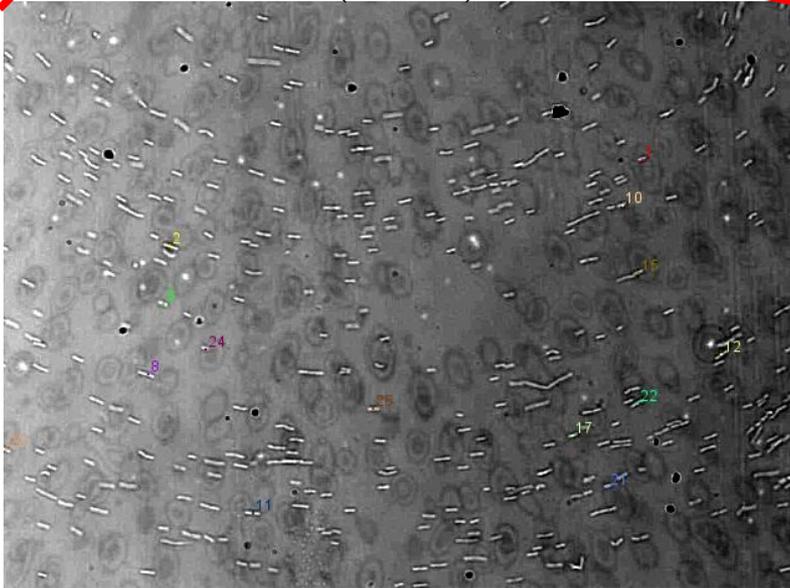
Number of bacteria swimming towards and away from the core is about the same at low c_B

Pure bend: Bacteria swim \parallel director; bipolar motion

$$c_B = 10^{14} / \text{m}^3$$

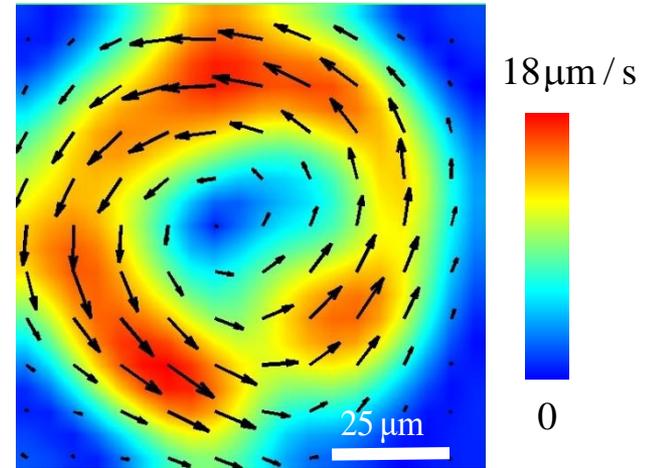
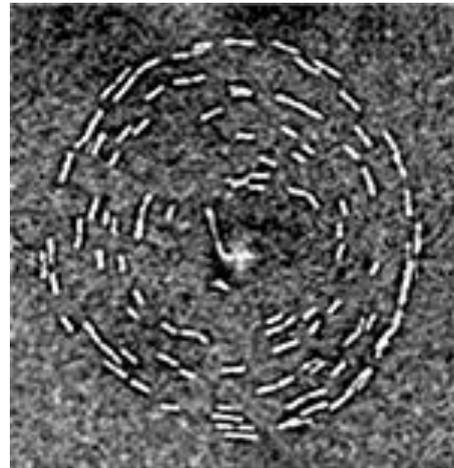
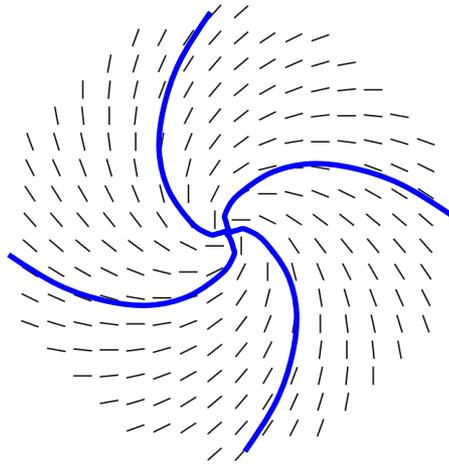


$$\text{bend} = \hat{\mathbf{n}} \times (\nabla \times \hat{\mathbf{n}})$$



Number of bacteria swimming to the right and to the left is the same, as expected

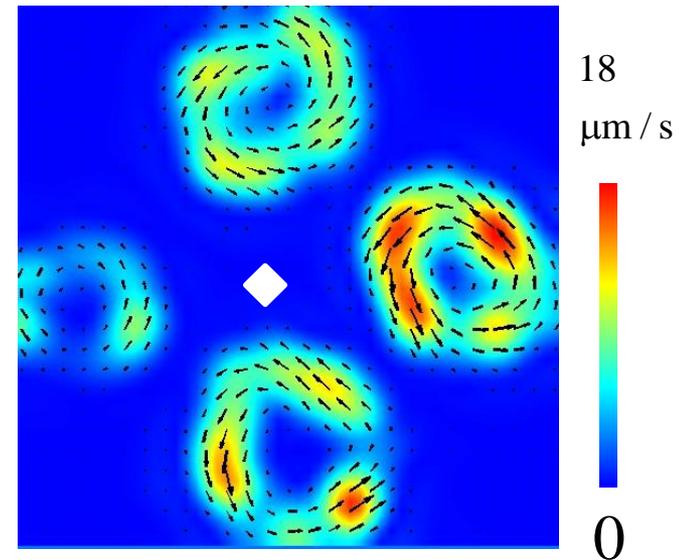
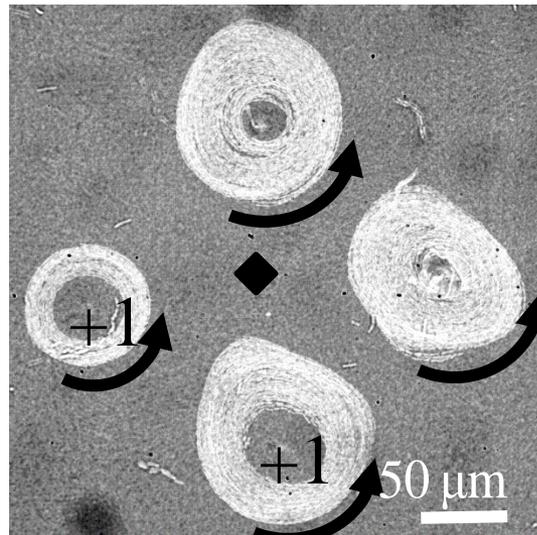
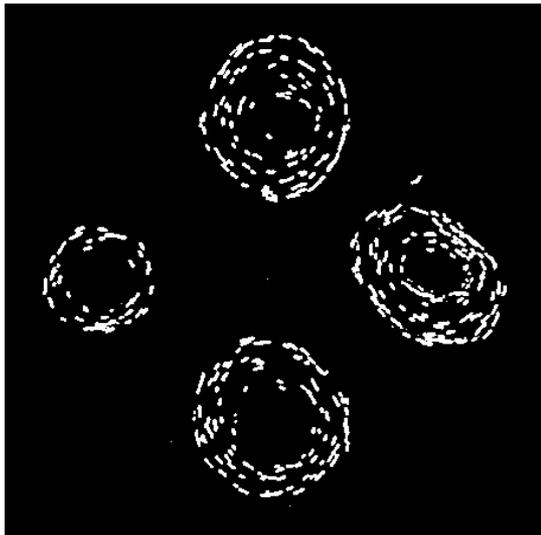
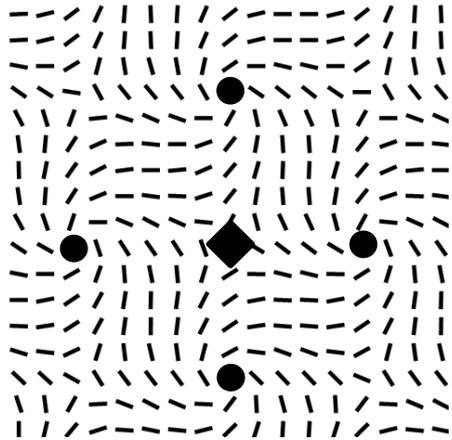
Mixed splay-bend: counterclockwise polar motion



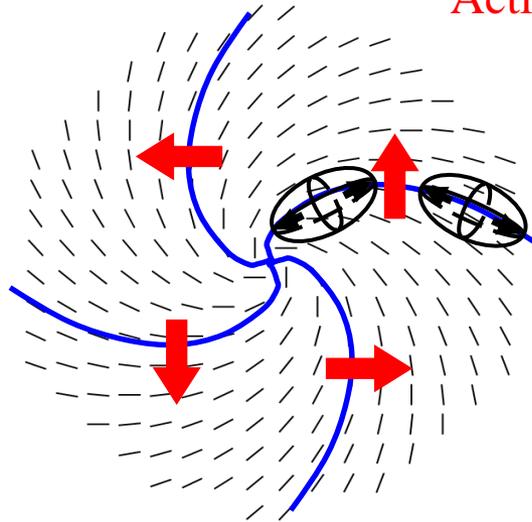
$$c_B = 10^{14} / \text{m}^3$$

Spiral vortex:
Counterclockwise collective
circular swimming; strictly polar

Periodic pattern of -1 and +1 defects: Bacteria gather and circulate around each +1, avoid -1



Mechanisms?



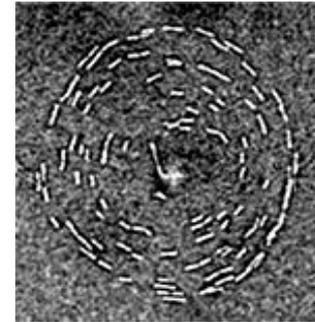
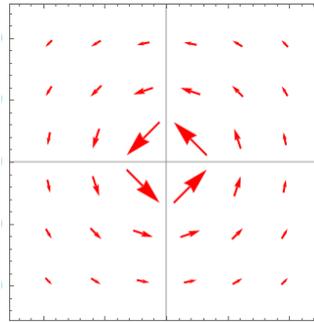
Active force

$$\mathbf{f} = \alpha \left[\hat{\mathbf{n}} \nabla \cdot \hat{\mathbf{n}} - \hat{\mathbf{n}} \times (\nabla \times \hat{\mathbf{n}}) \right]$$

$$\hat{\mathbf{n}} = (n_x, n_y, n_z) = (\cos \theta, \sin \theta, 0) \quad \theta(x, y) = \tan^{-1} \frac{y}{x} + \frac{\pi}{4}$$

$$\mathbf{f} = \{0, -\alpha / r\}$$

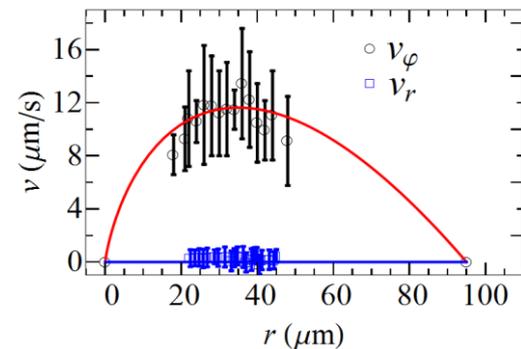
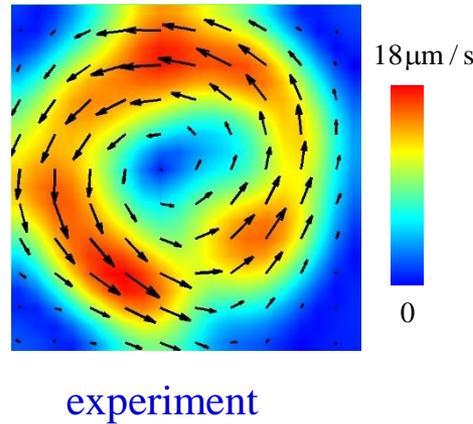
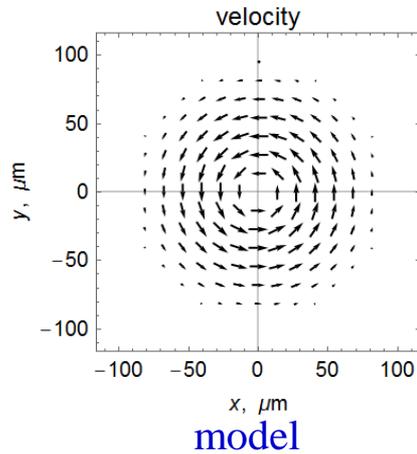
azimuthal force



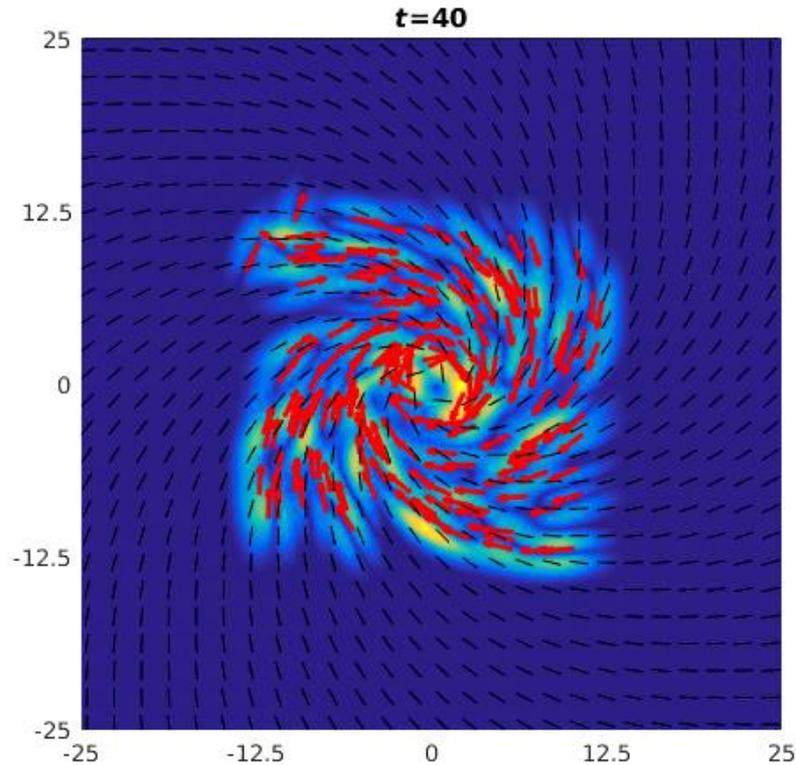
Opposing viscous force $\mathbf{f}_{drag} = \eta \nabla^2 \mathbf{v}$

Balance of drive and drag:

$$\mathbf{v} = \left\{ 0, \frac{\alpha r}{2\eta} \log \left(\frac{r}{r_0} \right) \right\}$$

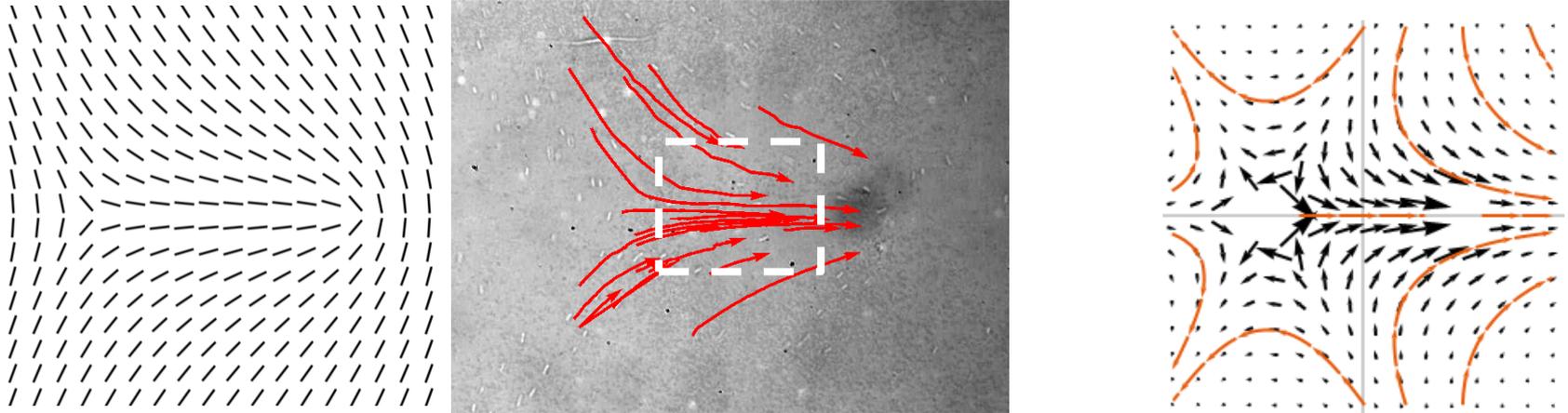


Mixed splay-bend: Forces unipolar circulation



Numerical simulations: Bacteria concentrate into toroidal swirls, concentration varies in space, I. Aranson, T. Turiv, R. Koizumi et al, submitted

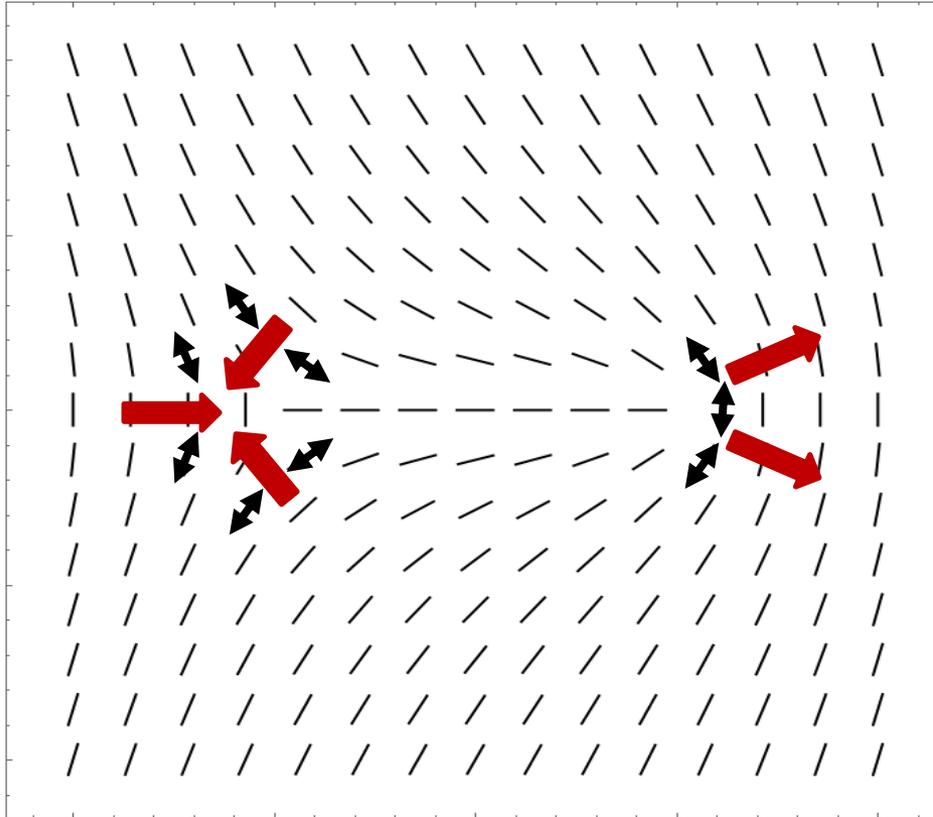
Pumping from -1/2 defect to +1/2 defect



Bacteria concentrate at the cores of +1/2 defects; avoid -1/2 defects

$$\mathbf{f} = \alpha \left[\hat{\mathbf{n}} \nabla \cdot \hat{\mathbf{n}} - \hat{\mathbf{n}} \times (\nabla \times \hat{\mathbf{n}}) \right]$$

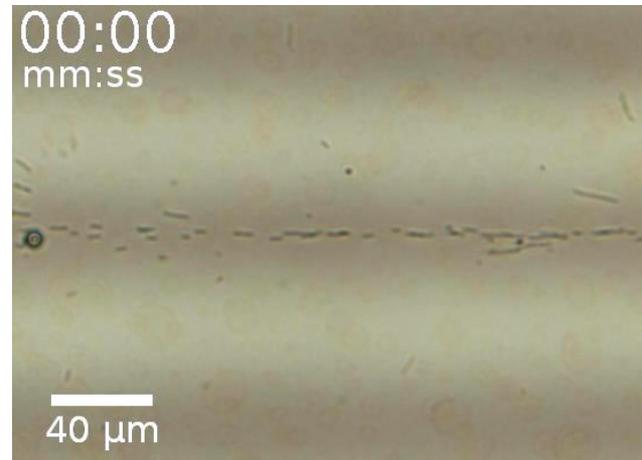
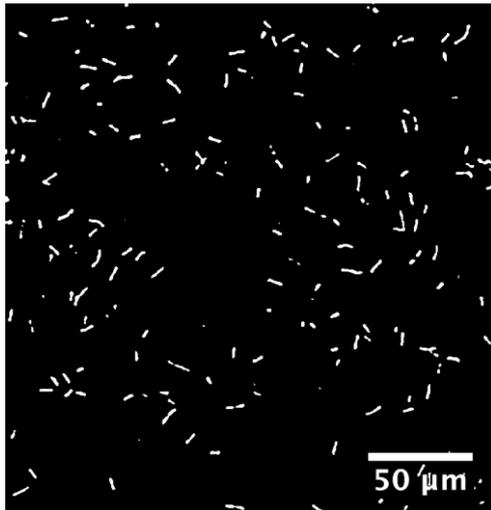
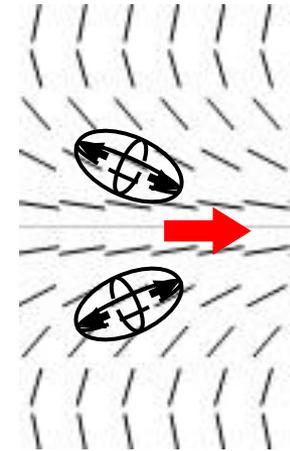
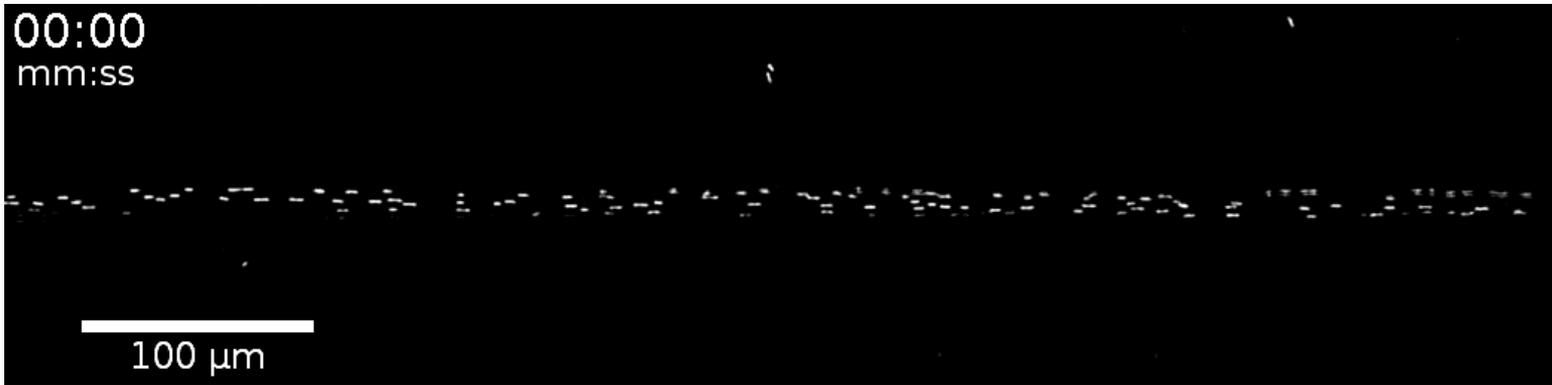
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$$\mathbf{f} = \alpha [\hat{\mathbf{n}} \nabla \cdot \hat{\mathbf{n}} - \hat{\mathbf{n}} \times (\nabla \times \hat{\mathbf{n}})]$$

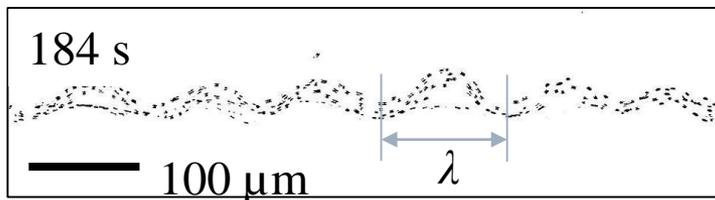
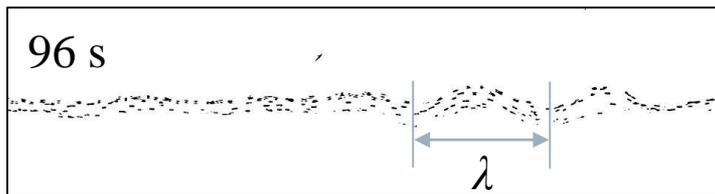
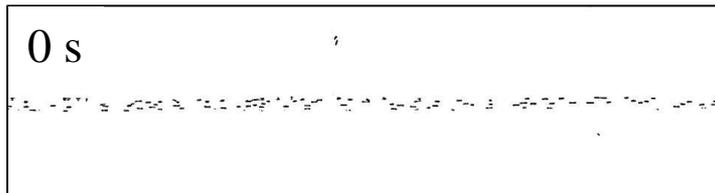
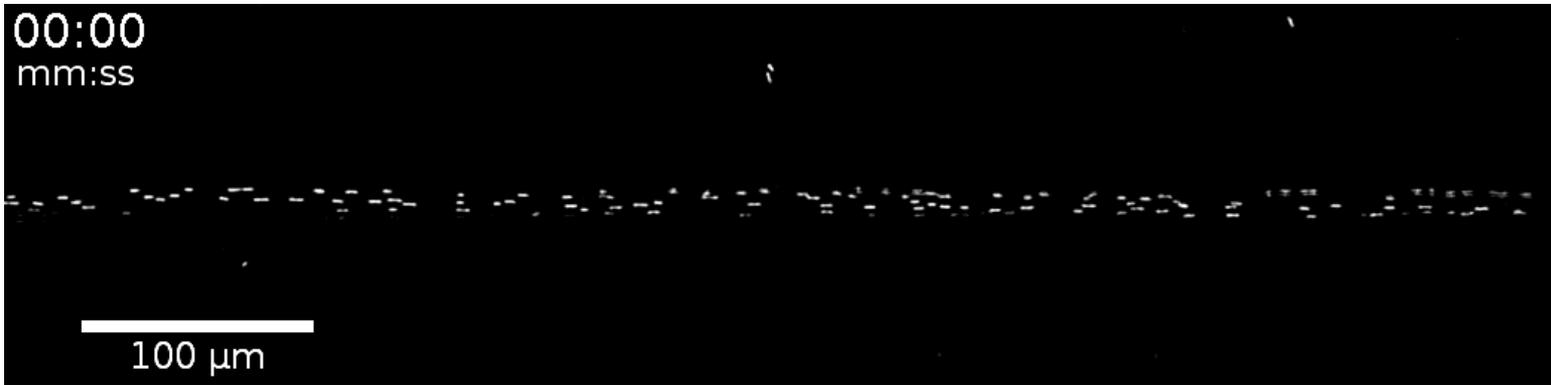
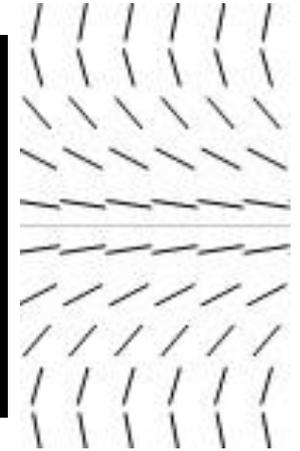
Activity tends to shift the +1/2 defect to the right-hand side

Unipolar swimming in concentrated jets:



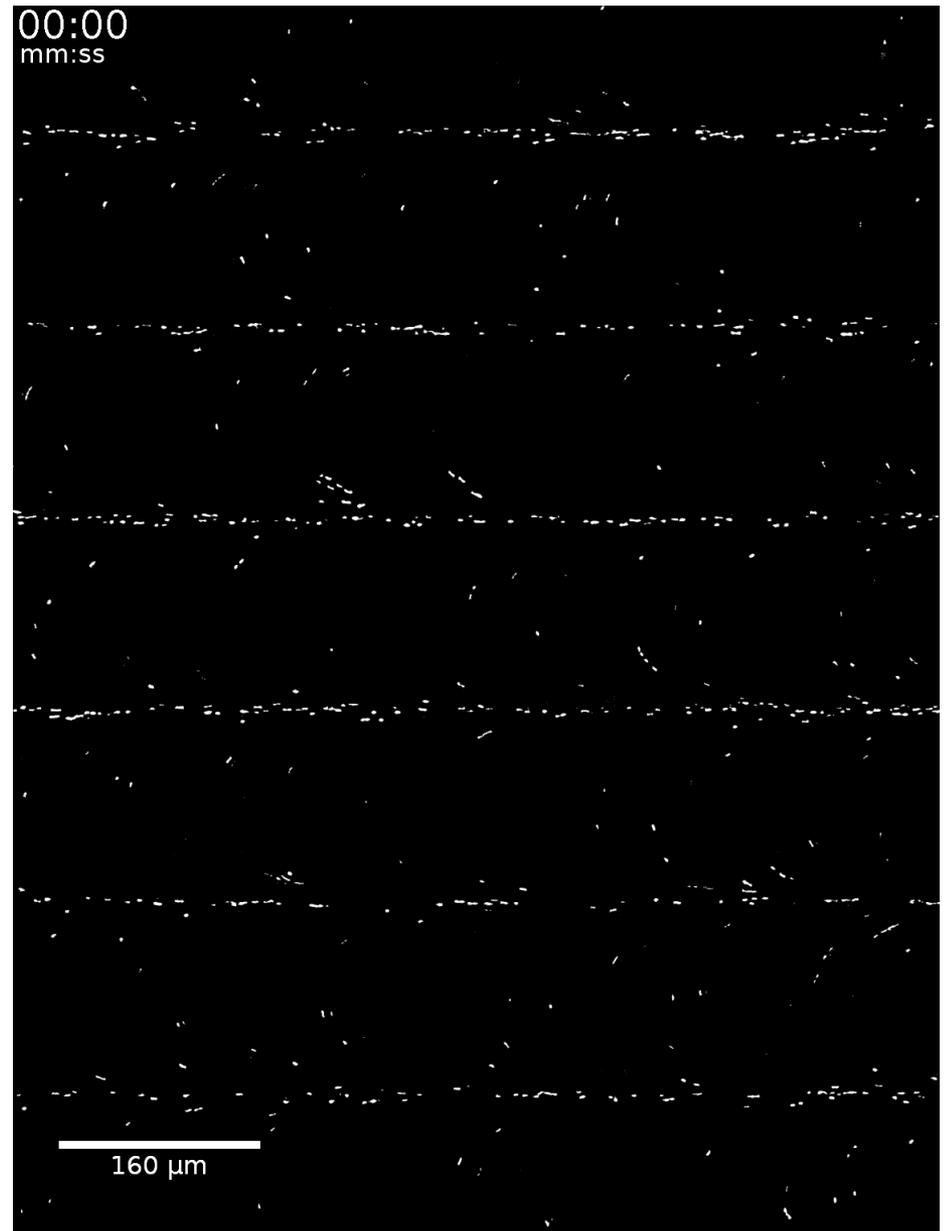
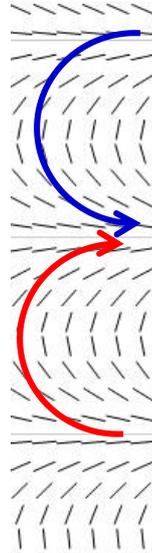
Isotropic phase; few degrees
higher temperature

Director bend limits undulations



As the jets undulate, the angle between the director and the trajectory increases; viscous drag increases and some bacteria are ejected; it helps to keep the period and amplitude of undulations constant; The jets are stable at $c < 7c_{\text{critical}}$ where c_{critical} is the critical concentration of instability in a uniform cell

Unipolar transport

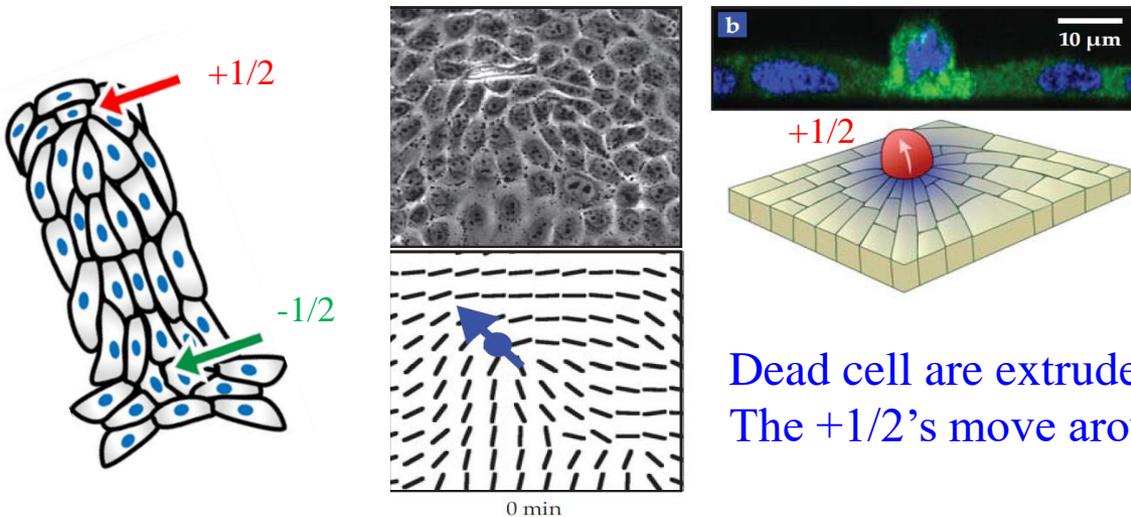


Conclusion-I:

- Bacteria in patterned nematic:
 - Patterns control trajectories, polarity and concentration of bacteria
 - Bacterial swirls and condensed jets are unidirectional and thus perform a useful work
 - Patterns cause unipolar threshold-less flow of bacteria, circular or linear
 - Concentration and thus activity are strongly non-uniform

Motivation No.2: Epithelium: Can we design it?

In epithelia, cells form an orientationally ordered nematic with topological defects, T.B. Saw et al, *Topological defects in epithelia govern cell death and extrusion*, Nature **544**, 212 (2017):



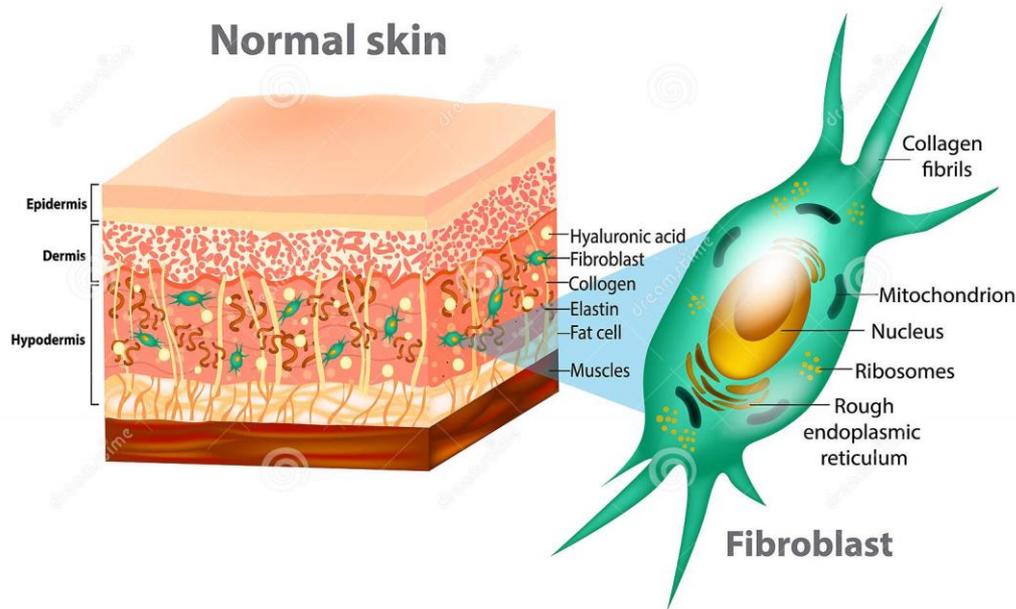
Dead cells are extruded at the $+1/2$ cores;
The $+1/2$'s move around in the plane of film

Topological defects are intrinsic to many other active matter systems

Sanchez et al, Nature **491**, 431 (2012), Zhou et al, PNAS 111, 1265 (2014)

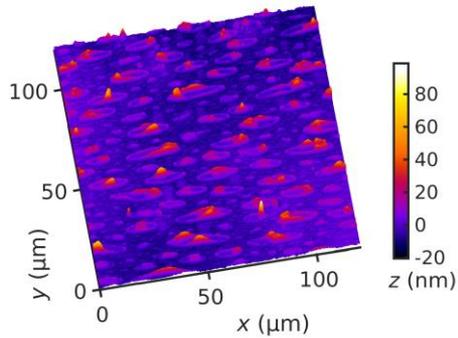
We do not know *where* the defects would emerge nor *how to control* them
Can we develop a template to produce and pin the defects at predefined locations?

Human dermal fibroblast cells

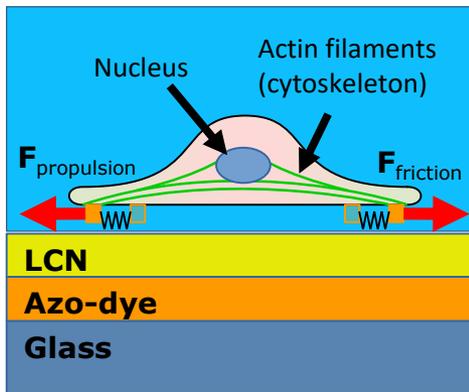
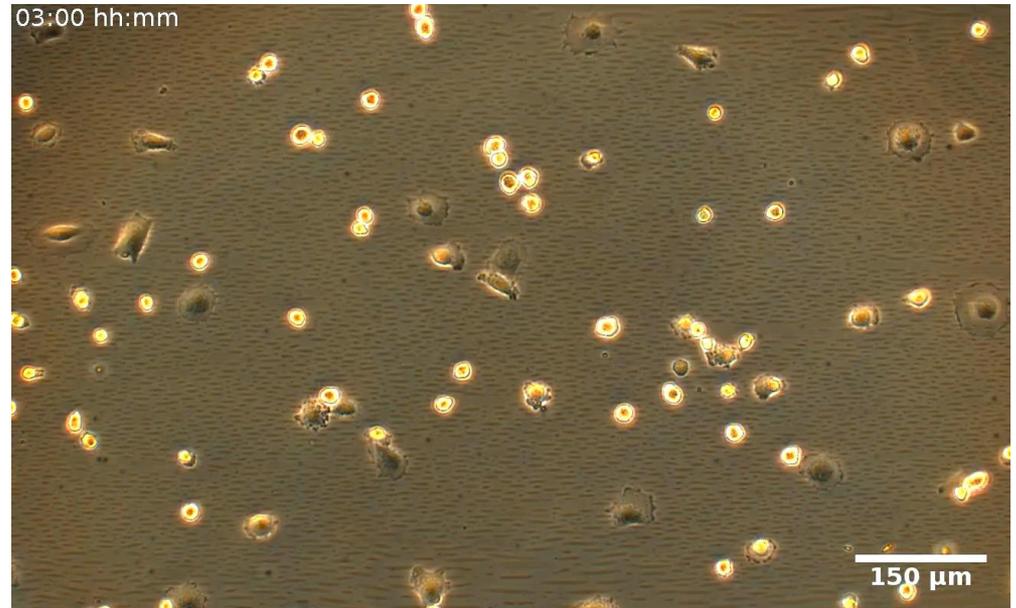


Fibroblast cells are part of skin responsible for generation of connective tissue and healing wounds

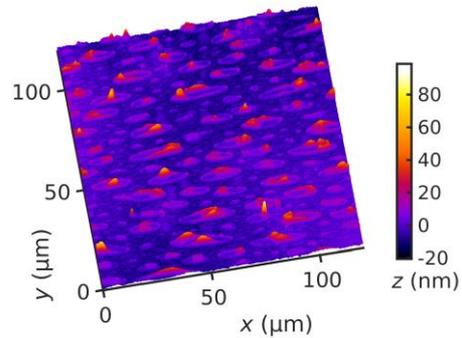
Deposition of cells onto LCN substrate from water dispersion



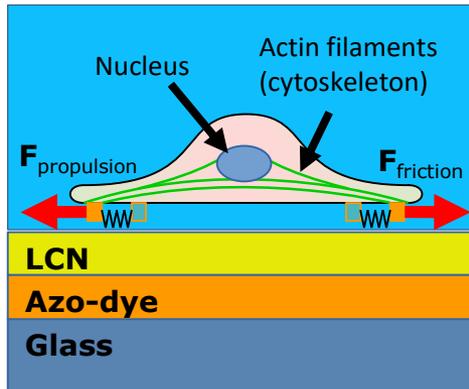
Seeding of human dermal fibroblast cells onto a uniformly aligned LCN substrate, top view



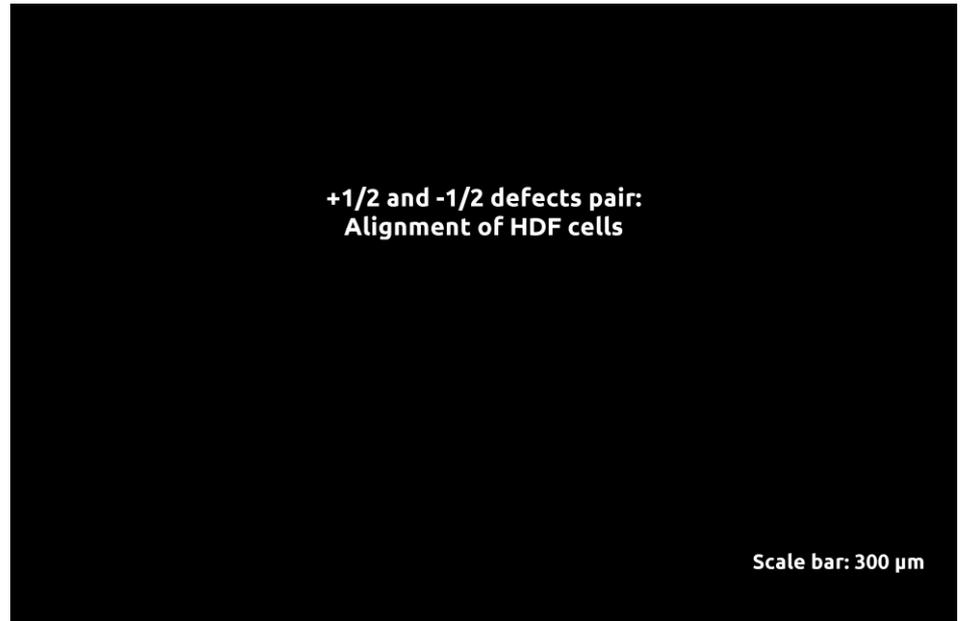
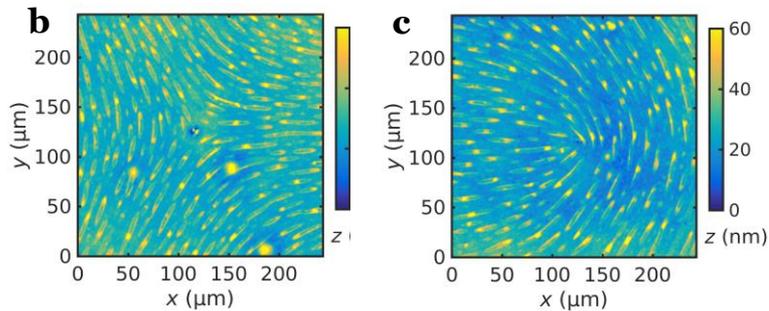
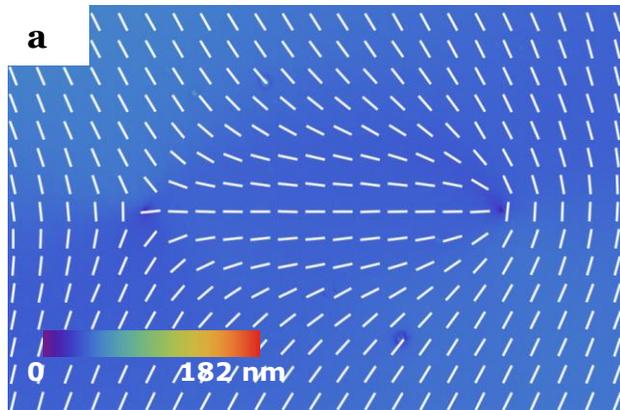
Alignment of human cells at uniform LCN substrate



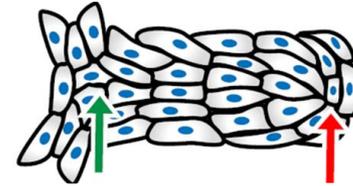
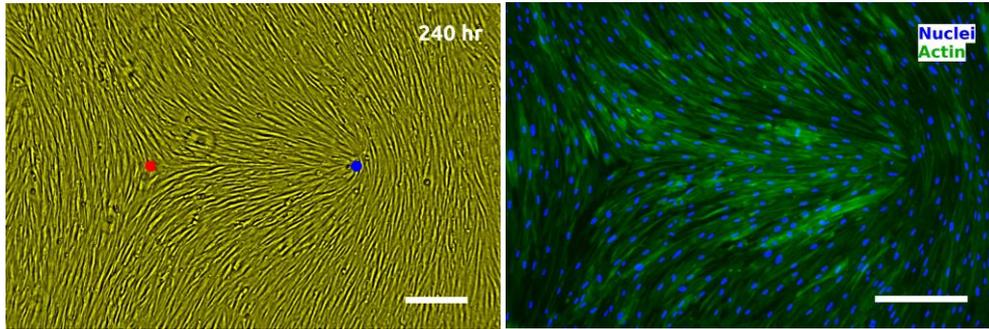
Uniform alignment of human dermal fibroblast cells at a uniform LCN substrate



Patterns with $+1/2$ and $-1/2$ topological defects

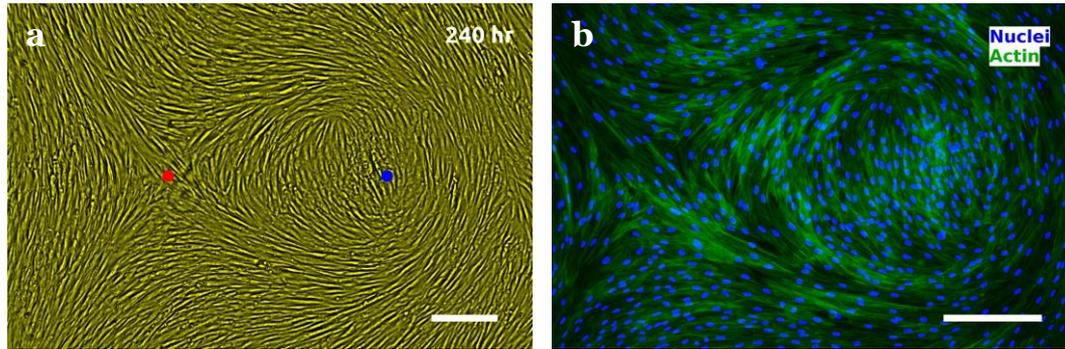


Patterns with $+1/2$ and $-1/2$ topological defects



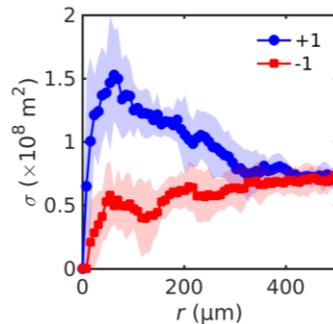
The templated tissue with $1/2$ and $-1/2$ defects closely resembles the natural epithelium tissue, with that difference that the defects do not leave the predesigned locations.

Patterns with +1 circular and -1 defects: Different concentration and phenotype of cells



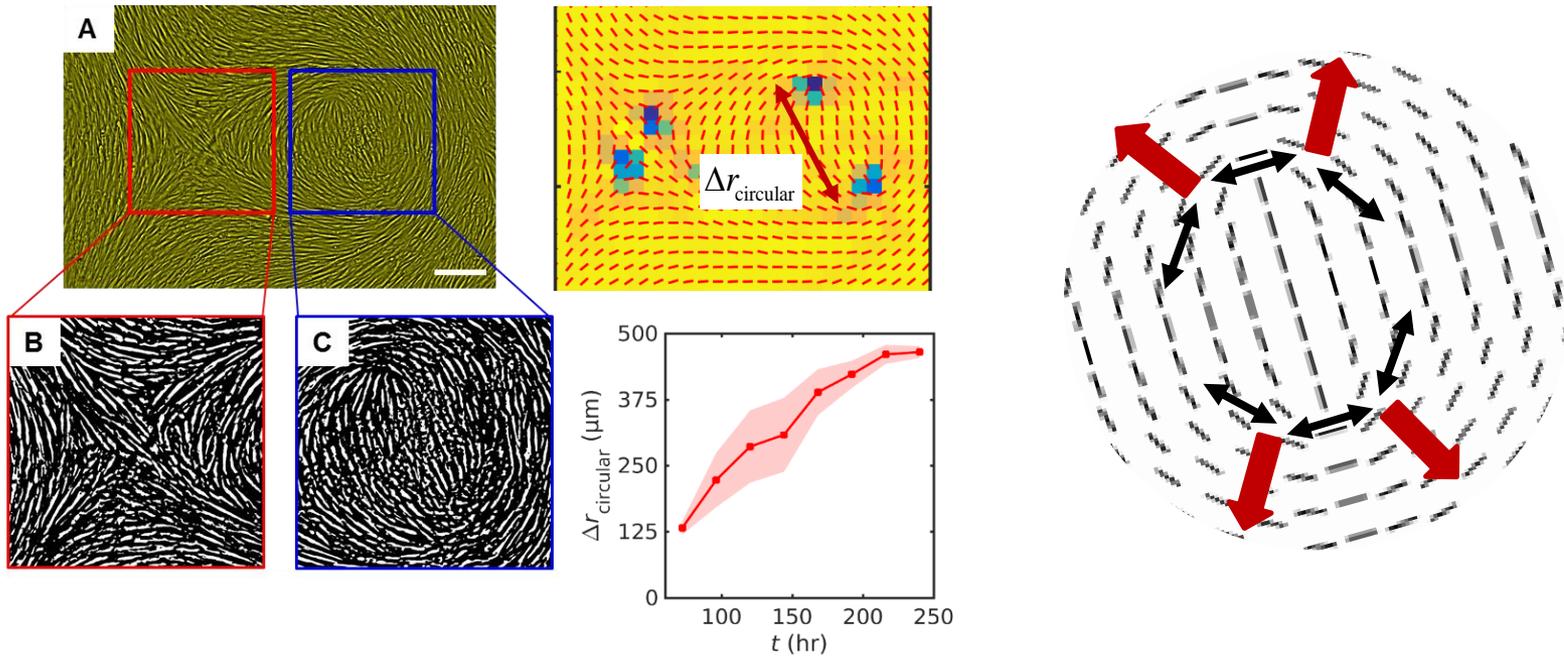
All scale bars are 300 μm

Density of cells
vs radial distance
from the core



+1 defect core attracts high concentration of cells while -1 core depletes them. Since the cells are in contact with each other, variation of density leads to variation in phenotype/shape: length/width ratio = 2.6 near +1 defects and = 5.8 near -1 defect.

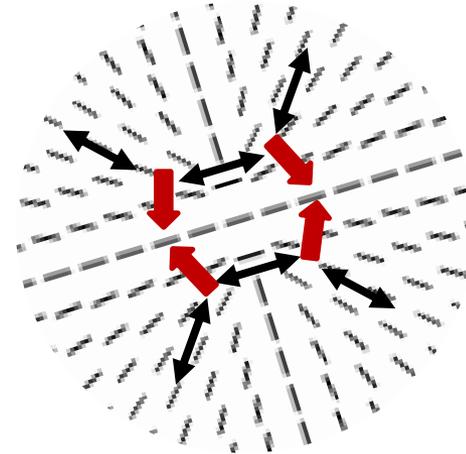
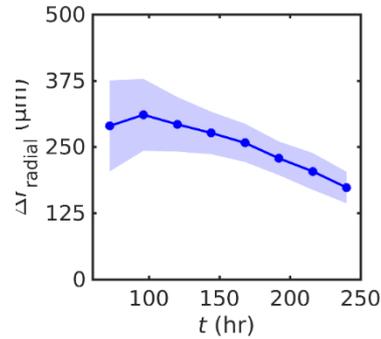
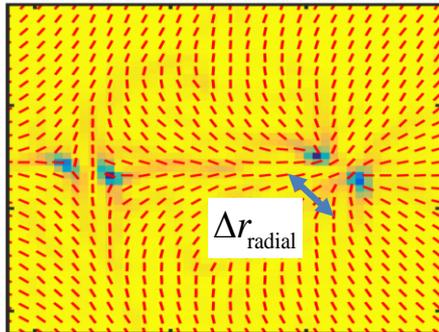
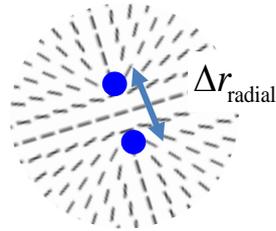
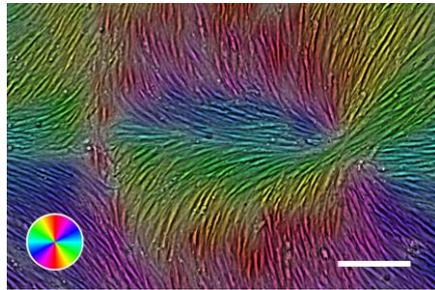
Core splitting of +1 circular defects into pairs of +1/2s



Circular +1 splits into +1/2 defects that move away from each other, which is expected as the elastic energy $\sim(\text{strength})^2$;

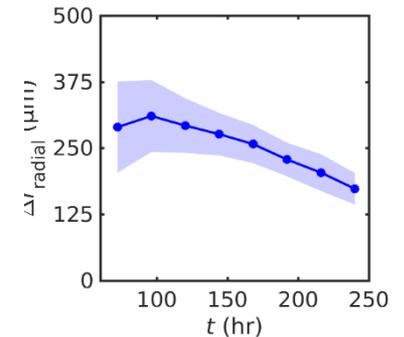
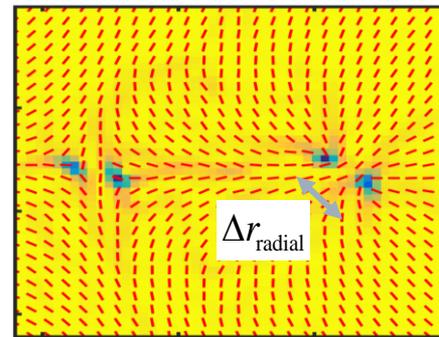
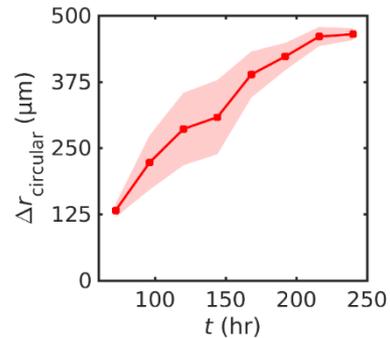
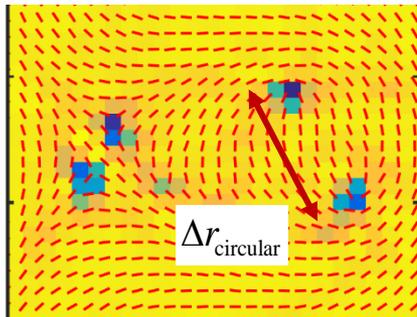
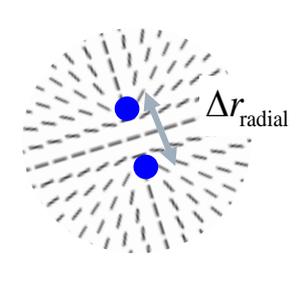
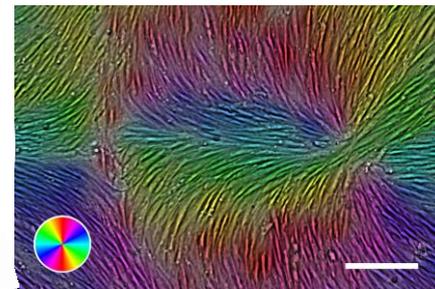
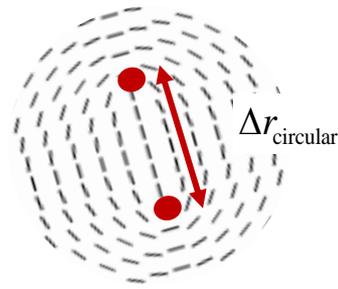
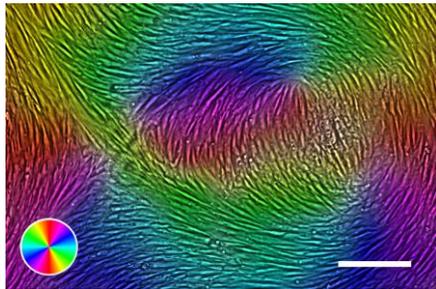
Note that the -1 defect splitting distance is shorter

Core splitting of +1 circular and radial defects into pairs of +1/2s



Radial +1 splits into +1/2 defects that move towards each other, defying elasticity!

Core splitting of +1 circular and radial defects into pairs of +1/2s



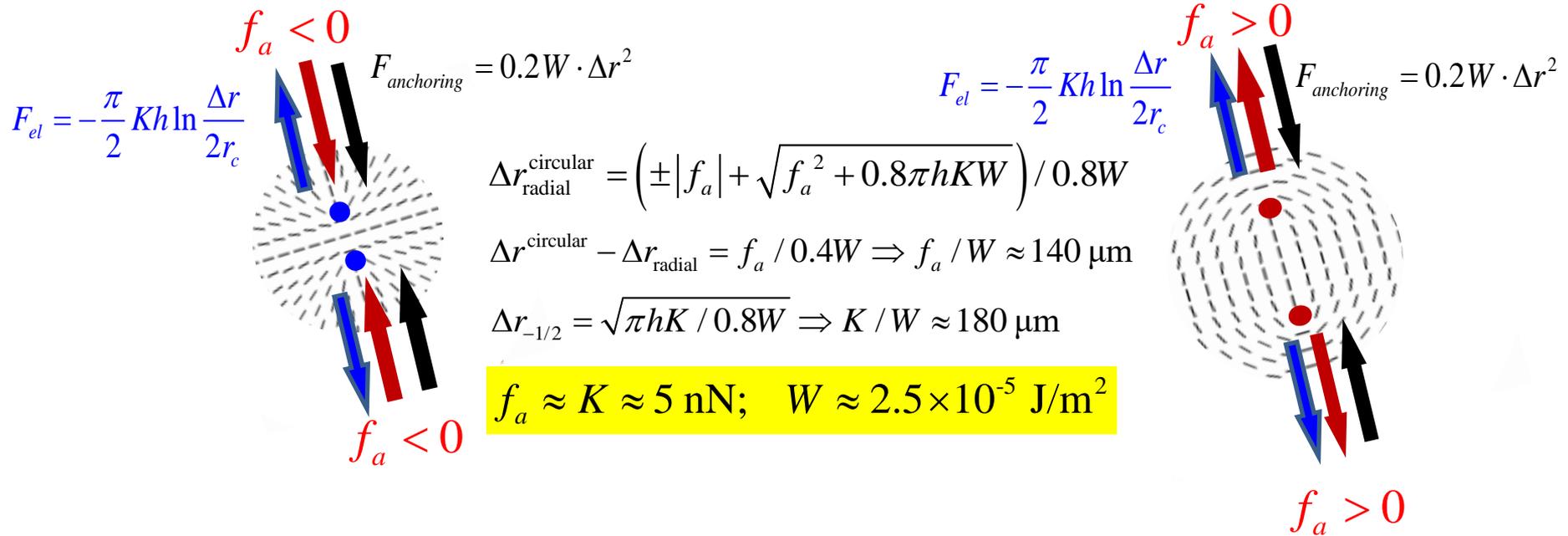
Circular +1 splits into +1/2 defects that move away from each other

Radial +1 splits into +1/2 defects that move towards each other, defying elasticity!

Side-by-side comparison of splitting of +1 circular and radial defects into pairs of 1/2 defects shows that the distance of splitting is influenced by active forces, in addition to elasticity and surface anchoring

Core splitting of +1 radial and circular defects into pairs of +1/2s

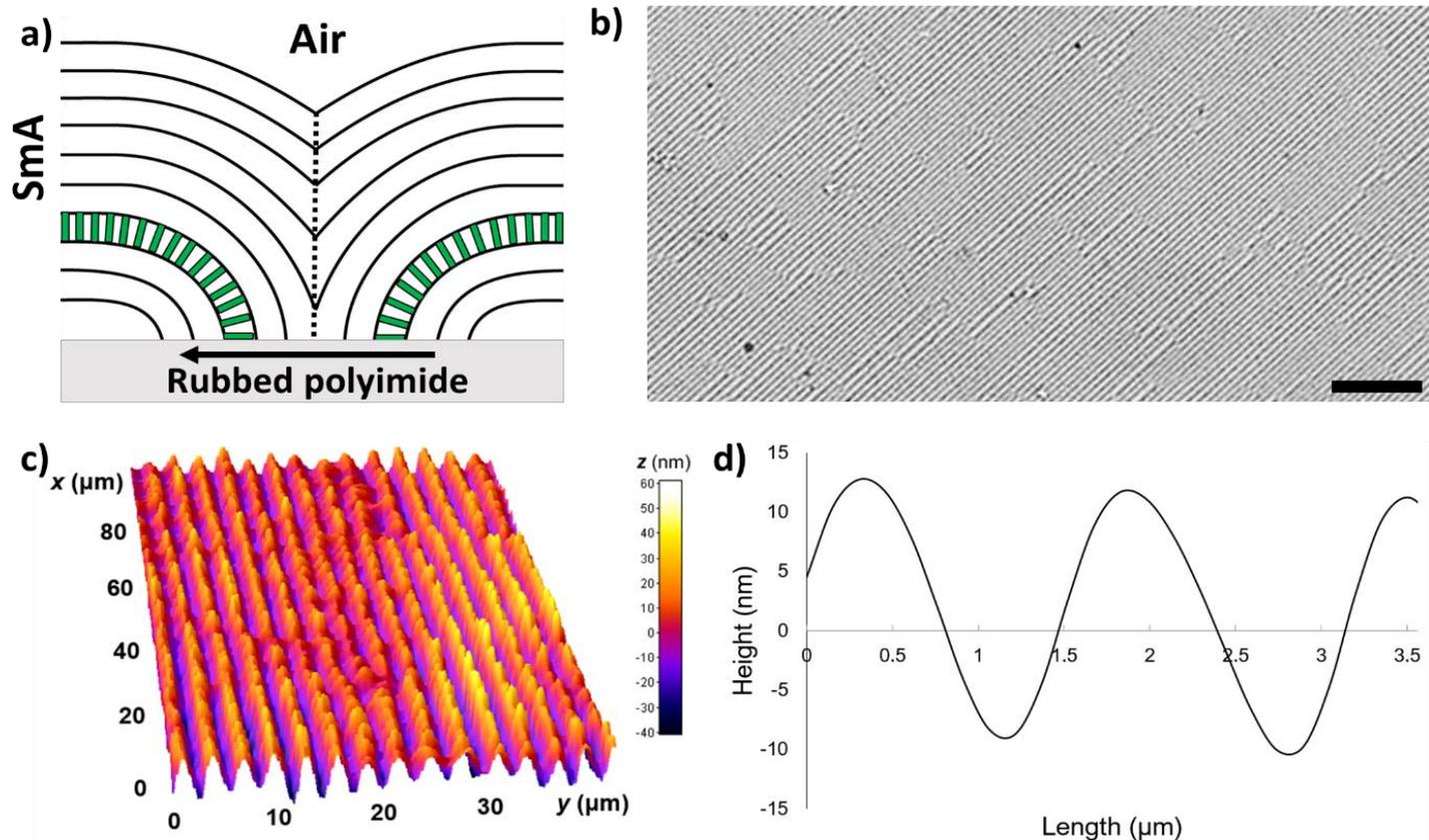
Balance of elasticity, surface anchoring and activity:



The experiment suggests that the cells are “pushers”; allows one to estimate tissue parameters such as the anchoring strength W , elastic modulus K and the active force f_a from the defect dynamics

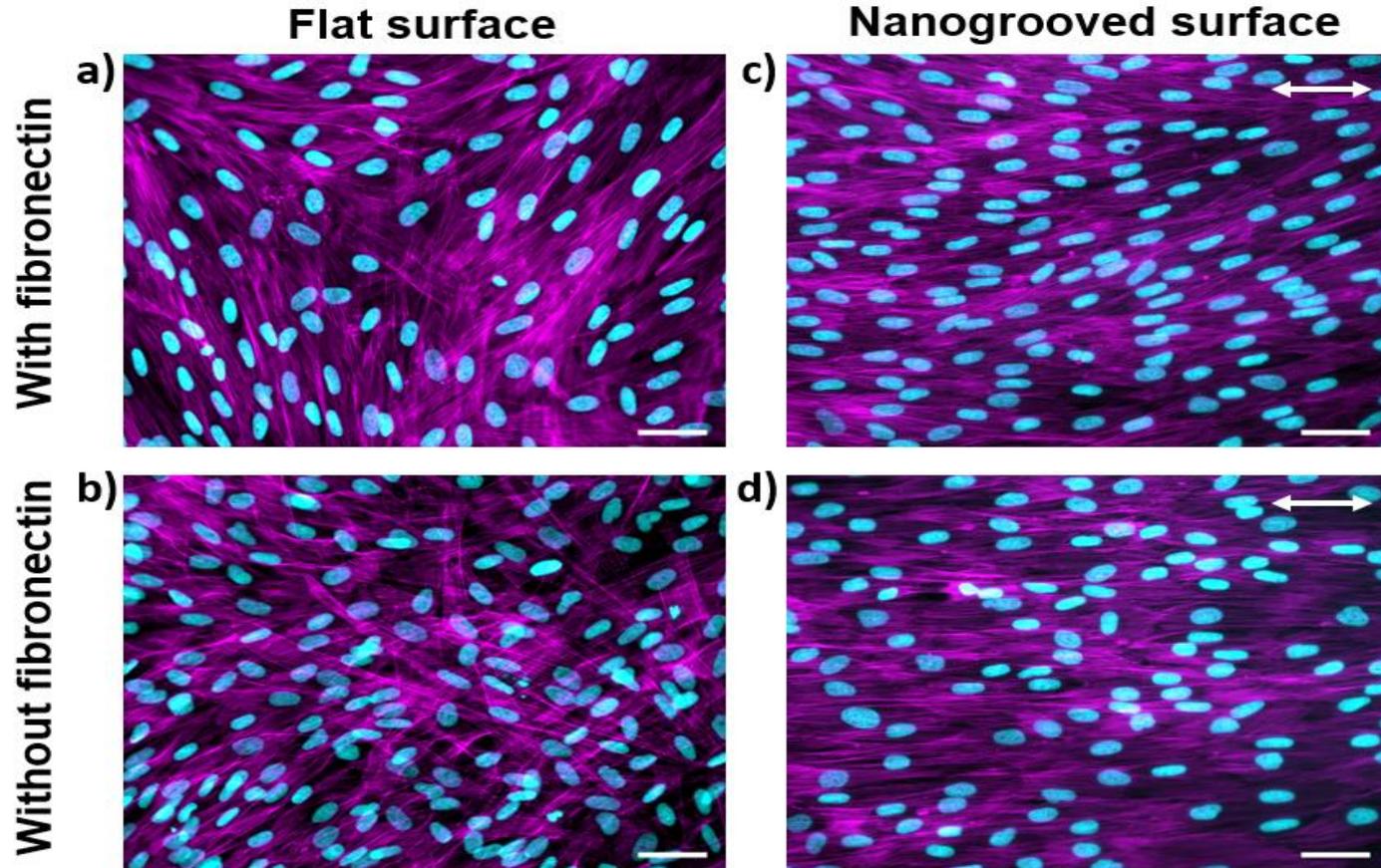
Alignment of human dermal fibroblast cells by smectic elastomers

A free surface of a smectic A develops undulations associated with the hybrid character of alignment and the requirement of layers equidistance (a); a mixture of a reactive monomer RM82, smectic A 8OCB and a photoinitiator Irgacure 651 can be polymerized in this undulated state (b,c) with nanogrooves amplitude reaching 25-30 nm (d)



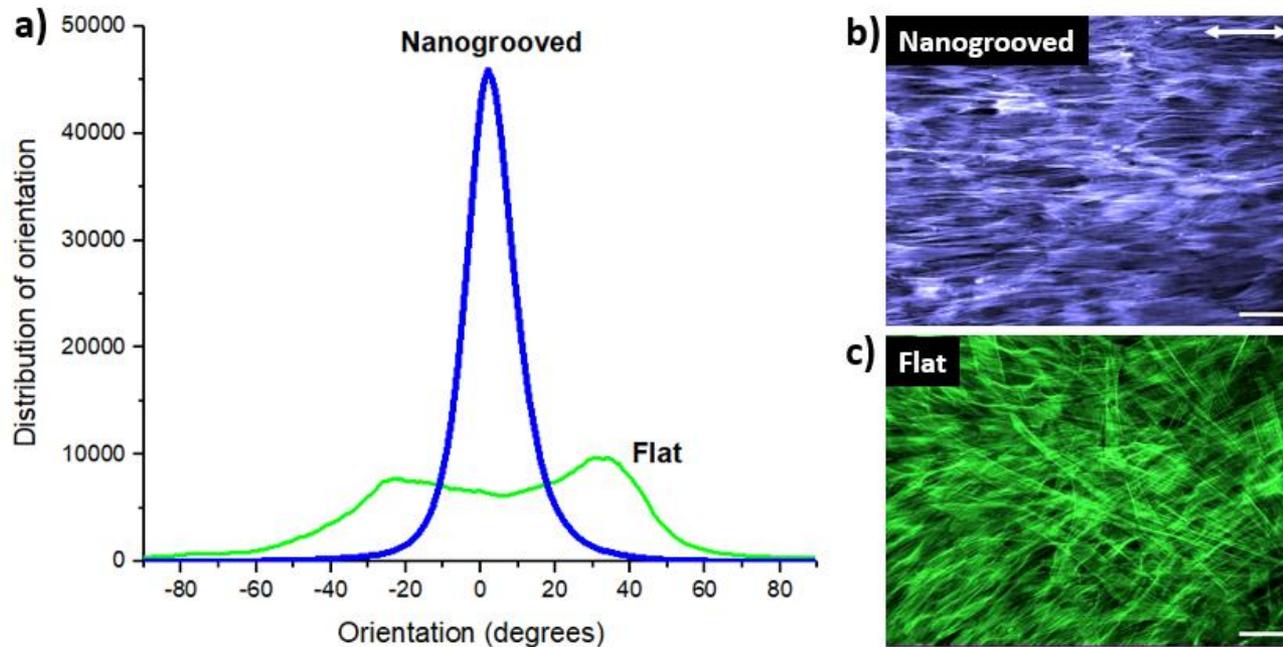
Alignment of human dermal fibroblast cells by smectic elastomers

Fluorescent microscopy images of hDF grown on substrate **a,b**) flat glass with fibronectin, **c,d**) glass with nanogrooved smectic A elastomer. Note the uniform alignment of cells by the smectic A elastomer nanogrooves oriented along the arrows. Scale bars 50 μm .



Alignment of human dermal fibroblast cells by smectic elastomers

Comparison of actin filament orientation by nanogrooved smectic A elastomer and by flat surface. Scale bars 50 μm .



Conclusion-II

- Liquid crystal elastomer coatings of nematic and smectic A type control human dermal fibroblast tissues
 - Patterned LC network in contact with aqueous solution produces elevations that guide patterned growth of cells
 - Undulation instability of smectic A elastomer creates nanogroves for cell alignment
 - Human dermal fibroblast tissues follow the predesigned LC network and behave as extensile active matter with $+1/2$ defects moving in the plane of the tissue
 - Patterns define the concentration and phenotype of cells
 - Difference in the splitting distance of the cores of radial and circular defects allows one to estimate the Frank elastic modulus of the tissue, surface anchoring and the active force

Summary

- Patterned liquid crystals can control trajectories, polarity of swimming and concentration distribution of swimming bacteria
- Patterned liquid crystal polymers can be used to design arrays of biological cells with predetermined locations of topological defects